

## **Radiation-Induced Effects In Lignin Model Compounds: A Pulse And Steady-State Radiolysis Study**

**Claudio A. Chuaqui<sup>1</sup>, Srinivasan Rajagopal<sup>1\*</sup>, Andras Kovács<sup>2</sup>, Terry Stepanik<sup>1</sup>,  
John Merritt<sup>1</sup>, István György<sup>2</sup>, Richard Whitehouse<sup>1</sup> and Don Ewing<sup>1</sup>**

<sup>1</sup>Research Chemistry Branch,  
AECL Research, Whiteshell Laboratories,  
Pinawa, Manitoba, Canada R0E 1L0

<sup>2</sup>Institute of Isotopes of the Hungarian Academy of Sciences,  
P.O. Box 77, H-1525 Budapest, Hungary

*(Received in USA 12 January 1993, accepted 18 August 1993)*

*Key Words* pulse radiolysis, gamma radiolysis, irradiation, lignin, model compounds

**Abstract** A lignin model compound  $\beta$ -(2-methoxyphenoxy)-3,4-dimethoxyacetophenone (**1**) was subjected to <sup>60</sup>Co gamma irradiation and pulse radiolysis under different conditions (dose, medium, pH). Interaction with hydroxyl radicals resulted in aryl hydroxylation, fragmentation, and cleavage. A hydroxylation-cleavage pathway was found at all pH values. At alkaline pH, fragmentation reactions were also observed. The predominant reaction with solvated electrons was fragmentation of the  $\beta$ -aryl ether bond followed by cleavage reactions.

### **INTRODUCTION**

Lignin, the second most abundant component in wood after cellulose, is produced in large amounts as a waste product in the pulping process<sup>1</sup>. In efforts to develop new industrial uses for ionizing radiation, the irradiation on wood, pulp, pulp mill

effluent and lignin is being examined in our laboratories. In pulping, pretreatment of wood with ionizing radiation resulted in numerous effects including a 20-25% net reduction in energy requirements<sup>2</sup>. Diverse effects were observed upon irradiation of both lignin isolated from irradiated wood, and commercial lignin samples<sup>3,4</sup>. Depending on the conditions used, polymerization, demethylation and degradative processes could coexist.

Because of the complex structure of lignin<sup>5</sup>, a lignin model compound  $\beta$ -(2-methoxyphenoxy)-3,4-dimethoxyacetophenone (compound I, Fig. 2, #13) was chosen for investigation of radiation-induced degradative pathways occurring in lignin. Such compounds have been used extensively in mechanistic studies<sup>6-8</sup> as their structures are representative of the arylglycerol- $\beta$ -aryl ether units that make up 30-50% of the repeating units in lignin<sup>6</sup>.

The results of irradiations performed on aqueous solutions of compound I are described here. Pulse radiolysis was used to monitor formation of transient species, while HPLC was used to identify stable radiolytic products.

## 1. RESULTS AND DISCUSSION

Reaction pathways are described using the convention initiated by Dimmel<sup>9</sup>. "Fragmentation" refers to cleavage of the  $\beta$ -aryl ether bond. Decomposition, degradation, dehydration and dehydrogenation denote breakage of bonds other than the  $\beta$ -aryl ether bond and are grouped under "Cleavage" reactions. "Hydroxylation" relates to an hydroxyl adduct on an aromatic ring as a  $\sigma$ - or  $\pi$ -complex. "Combination" relates to polymerization, dimerization and disproportionation reactions. "Demethoxylation" corresponds to ipso substitution of a methoxyl group by a hydroxyl group.

### 1.1 Radiolysis Under Oxidative Conditions

In  $N_2O$ -saturated water, the dominant path to oxidative radiolysis is via hydroxyl radicals<sup>10</sup>. Hydroxyl radicals are strong oxidizing species known to abstract hydrogen from aliphatic hydrocarbons<sup>11</sup>. With aromatics, the major reaction is addition of hydroxyl radical to the aromatic ring to form hydroxyl adducts<sup>10</sup>.

**1.1.1 Steady State Radiolysis** Thirteen major peaks were found upon HPLC of I (0.1 mM in  $H_2O$ ) irradiated under oxidizing conditions (Fig. 1A). The identified products (Fig. 2) could be grouped into 3 sets based on the reactions assumed to be involved in their formation. The first set is the hydroxylated and demethoxylated products in peaks 10, 11, and 12. The second set is the fragmentation products 8 and 9, while the third set contains the more polar cleavage products, 1-7.

At pH 3, compound I disintegrated almost linearly between 0.03-1 kGy and exponentially thereafter (Fig. 3). The hydroxylated products (peaks 10, 11, & 12) and the fragmentation products guaiacol (8), and dimethoxyacetophenone (9, DMAP) were formed in low concentrations at 0.03 kGy, increased with increasing dose, then decreased at higher doses as they underwent secondary radiolysis. Dimethoxyphenol, DMP, increased with dose to 0.5 kGy, then declined. Cleavage products 1-7 appeared as primary products, even at the lowest dose measured.

The steady-state radiolysis pattern of I at pH 7.2 differed from that at pH 3.0 (results not shown). At pH 7.2, I radiolysed easier, with approximately 10% remaining after a dose of 1 kGy. The hydroxylated products (10, 11, 12) reached their maxima at 0.5 kGy, whereas the profile for DMP remained the same as at pH 3. Fragmentation products, guaiacol and DMAP showed increased concentrations at lower doses, indicating increased fragmentation at pH 7.2.

At pH 11, almost 75% of I was radiolysed at 0.5 kGy. Hydroxylation products found in peaks 10, 11 & 12 showed similar profiles with sharp declines beyond 0.5 kGy. Guaiacol and DMAP formed in significantly higher amounts indicating enhanced fragmentation. However, DMP formed in very low concentrations compared to at lower pH.

**1.1.2 Pulse Radiolysis** The light absorption spectrum of species formed upon pulse radiolysis of I (0.1 mM in H<sub>2</sub>O) taken immediately after the 2.6  $\mu$ s electron pulse gave maxima at 350 and 420 nm (Fig. 4). Similar absorption spectra were observed at acidic pH. Such "double peak" spectra appearing quickly after the pulse have been related to the formation of hydroxycyclohexadienyl radicals (hydroxyl adducts)<sup>10,12</sup>. Thus the absorption spectrum for I is attributed to the instant formation of the hydroxyl adduct. The rapid decay of both absorbing species (Table 1) by a second-order process in the microsecond time scale, is also characteristic of bimolecular radical-radical reactions of hydroxyl adducts<sup>13</sup>.

A "long-lived" remaining absorption, which was more prevalent at higher dose per pulse values (i.e., 50 Gy/pulse), was also observed at  $\sim$ 350 nm. This absorption appeared a few hundred microseconds after the electron pulse (Fig. 4).

The optical absorption spectra measured at pH 3.0, 7.2, and 11.0, were similar with the exception that at pH 11, the long-lived absorption at 350 nm was more intense. This absorption was stable even in the millisecond time scale. In addition, a small absorption was observed in the 370–400 nm range at pH 11. This species was attributed to phenoxyl radical formation in small yield resulting from oxidative displacement of methoxyl groups via ipso attack of hydroxyl radical<sup>14</sup>. This demethoxylation pathway has also been found on pulse radiolysis of anisole, (results not shown) where the absorbance of the phenoxyl radical continued to increase long after the hydroxyl adduct had decayed.

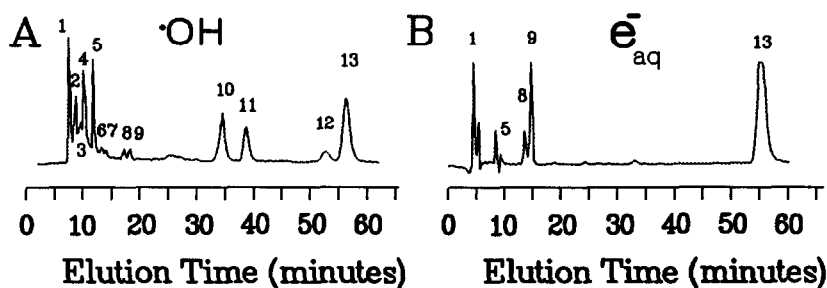


Fig. 1 HPLC of I irradiated under oxidizing (A) and reducing (B) conditions at pH 3 and 1.0 kGy

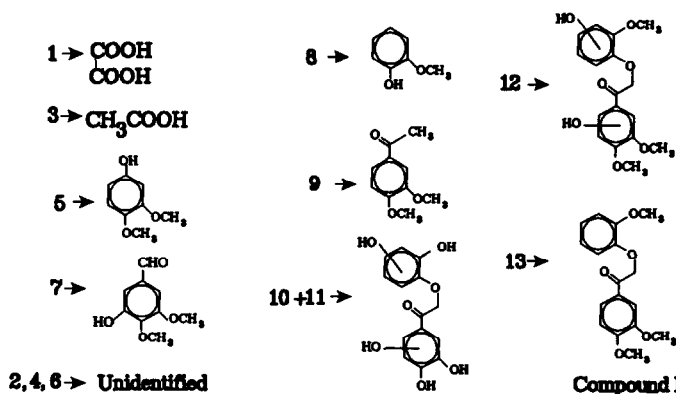


Fig. 2 Characterization of major radiolysis products by HPLC and <sup>1</sup>H NMR

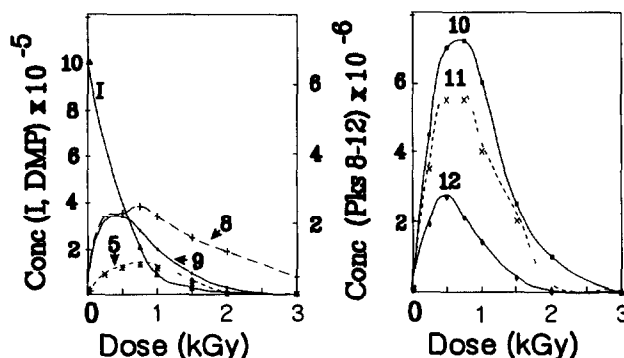


Fig. 3 Area profile of major radiolytic products as a function of dose

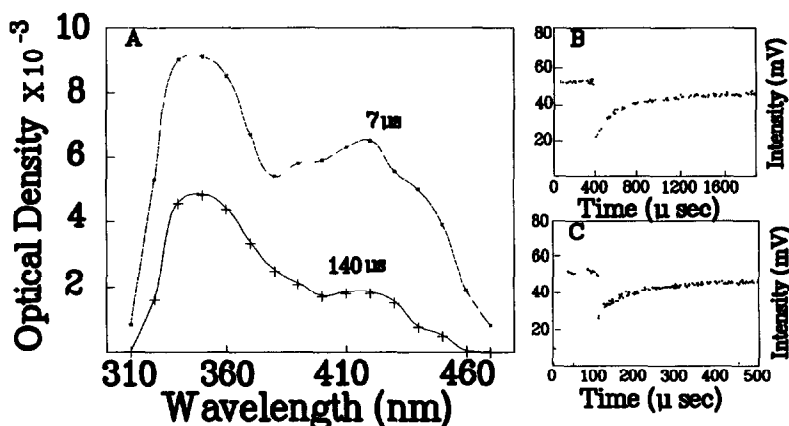


Fig. 4 (A) Optical absorption spectra of the hydroxyl adduct of I taken 7  $\mu$ s and 140  $\mu$ s after the 2.6  $\mu$ s electron pulse (pH 7.2), (B) Oscilloscope traces related to the decay of the absorbing species observed at 350 nm, and (C) at 420 nm.

**1.1.3 Summary of Radiolysis Under Oxidative Conditions** The pulse radiolysis results support the instant formation of hydroxyl adducts of I. Phenoxyl radicals are also formed in small amounts presumably via ipso hydroxylation. These results agree with steady-state data which show that hydroxyl radical-mediated radiolysis of I occurs rather selectively.

Hydroxylated and demethoxylated products (10, 11, 12) predominated at all pH values. Significant fragmentation to 8, and 9 occurred only at higher pH indicating that abstraction of the methylene hydrogen proceeds more easily in alkaline medium. Cleavage to form 1-7 was observed at all pH values. Based on these results, the pathways shown in Fig. 5 are postulated.

At lower pH the hydroxyl radical preferentially attacks the aromatic rings of I. Hydroxylation occurs via formation of a  $\pi$ -adduct on the aromatic ring(s), to form products found in peak 12. These compounds probably result from primary reactions, since they form and disappear at low doses. The presence of hydroxylation products (in peaks 10 and 11) in high concentrations at all pH values supports a pathway involving ipso hydroxylation. Because the methoxyl group may direct the

attacking hydroxyl radical preferentially into the ortho and para positions on each ring, different hydroxylated products would be expected. Ipso hydroxylation can also occur at the aromatic carbons linked to the aliphatic bridge leading to fragmentation. The presence of DMP at pH 3 and 7.2 is indicative of ipso hydroxylation at the bridging carbon of the DMAP ring.

Table 1 Spectral Characteristics and Decay Rate Constants of Hydroxyl Adducts Formed in Compound I

pH	$\lambda_{\max}, \text{nm}$	$G_{\epsilon}$ (350 nm)	$G_{\epsilon}$ (420 nm)	$2k/\epsilon, \text{cm s}^{-1}$ (350 nm)	$2k/\epsilon, \text{cm s}^{-1}$ (420 nm)
3.01	345	$7.6 \times 10^3$	$5.9 \times 10^3$	$1.2 \times 10^6$	$1.3 \times 10^6$
7.20	350	$6.5 \times 10^3$	$4.4 \times 10^3$	$1.6 \times 10^6$	$1.5 \times 10^6$
11.0	350	$8.4 \times 10^3$	$5.0 \times 10^3$	-	$4.0 \times 10^6$

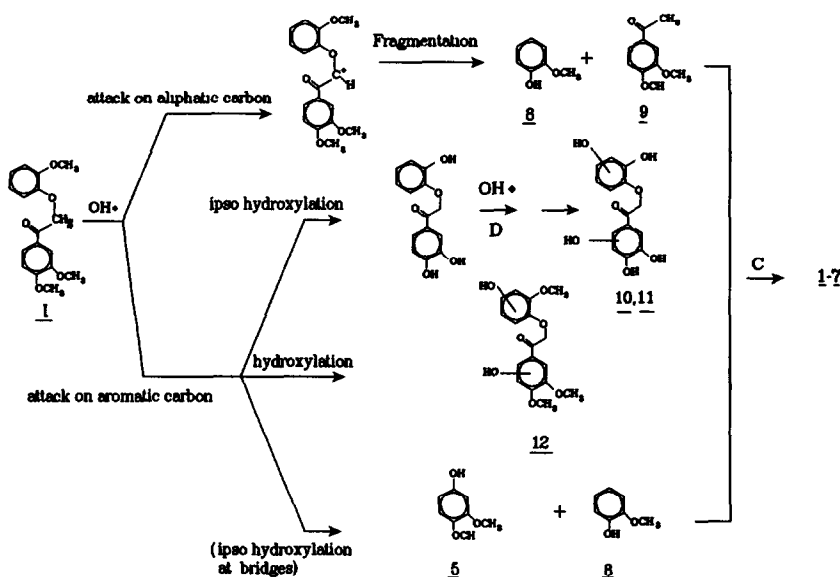


Fig. 5 Possible mechanistic pathways for radiolysis of I under  $\text{N}_2\text{O}$

Hydroxyl radical may also attack the aliphatic methylene carbon at high pH forming 8 and 9, although other possibilities exist, i.e.  $\text{O}^-$  formed via reaction 1, may abstract the methylene proton from I causing fragmentation, as the  $\text{pK}_a$  for this



reaction is 11.9<sup>15</sup>. Increased amounts of 8 and 9 at pH 11.0 imply the pH effect contributes to this mode of fragmentation.

## 1.2 Radiolysis Under Reducing Conditions

Electrons, one of the major transient products in radiolysis of water, are quickly solvated forming the strongly reducing species,  $e_{aq}^-$ . This species forms either anions or free radicals by bond scission reactions with substrates. A very selective reaction occurs with aromatic ketones, the adherence of electrons to the carbonyl group at diffusion controlled rates<sup>16</sup>. As shown below, I undergoes very specific reactions under reducing conditions.

**1.2.1 Steady State Radiolysis.** Fewer radiolytic products were formed under reducing conditions (Fig. 1B) compared to oxidizing conditions. The major products were the fragmentation products guaiacol (8) and dimethoxyacetophenone (9, DMAP). I decomposed almost linearly between 0-1 kGy (Fig. 6). Guaiacol (8) and DMAP (9) formed in higher concentration than under oxidizing conditions, increased with dose to 2 kGy then declined. DMP (5) decreased beyond 1 kGy. No hydroxylated derivatives of I were found.

Compound I fragmented much faster at higher pH. At pH 7.2 fragmentation reactions yielded guaiacol and DMAP. The DMP concentration remained stable up to 1.5 kGy. At pH 11, guaiacol and DMAP increased in concentration, while DMP was absent, possibly due to faster degradation through a combination of pH and radiation effects.

**1.2.2 Pulse Radiolysis.** The transient optical absorption spectrum of I varied little with pH. At pH 4, the spectrum taken immediately after the 2.6  $\mu$ s electron pulse had maxima at 350, 365, 385, and 420 nm. The maxima at 350 nm and 420 nm are characteristic of ketyl radicals formed by reaction of  $e_{aq}^-$  with aromatic carbonyl compounds. These species formed rapidly after the pulse and decayed by second-order kinetics (Table 2) in agreement with data by Hayon *et al*.<sup>16</sup>

The similarity in absorption spectra at pH 4 and 13 implies that the ketyl radical anion does not form at pH 13. For example we found different spectra for anisoin at pH 3 and 11 (results not shown). The different spectra were assumed to be due to the ketyl radical and the ketyl radical anion of anisoin. Similar results were found by Hayon *et al*.<sup>16</sup>

The maxima at 365 and 385 nm are similar to that found upon pulse radiolysis of guaiacol in  $N_2O$ -saturated solution (results not shown). With guaiacol, this absorption was probably due to the phenoxyl radical, since a very similar absorption was found when  $NaN_3$  (used to oxidize aromatic compounds to phenoxyl radicals) was included.

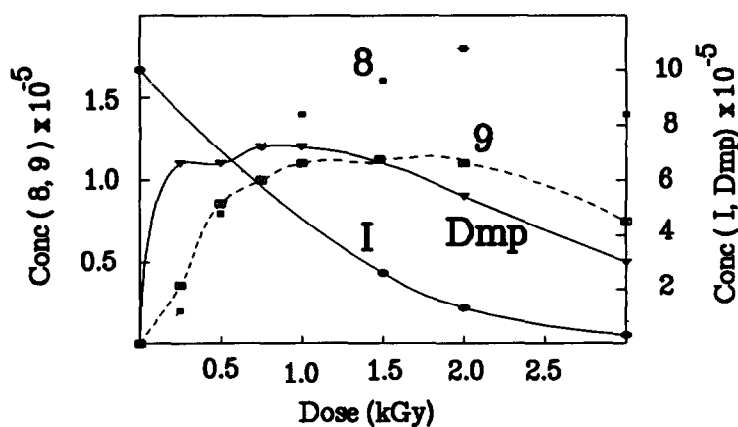


Fig. 6 Area profile of major components from I irradiated under reducing conditions



containing a hydroxyl group at the para position of the arylglycerol moiety were found to be much more susceptible to acid hydrolysis than analogs containing a methoxyl group

Ciaramitaro and Steelink<sup>18</sup> studied the acid hydrolysis of 4-hydroxybenzylphenyl ethers and their acetates (to mimic ester structural linkages in lignin). The acetates required 24 h for fragmentation whereas the free phenols were completely hydrolysed in 8 h. Sarkanen and Hoo<sup>19</sup> determined a relative rate of 4:1 for fragmentation of the  $\beta$ -aryl ether bond in a phenolic model compared to its methoxyl analog. Several cases in the literature support this fact<sup>19,20</sup>

Thus non-phenolic lignin model compounds appear to be much less reactive to chemical fragmentation. In this context, it is interesting that **I** fragments very easily under ionizing radiation, especially in the presence of solvated electrons. As non-phenolic units containing  $\alpha$ - and  $\beta$ -aryl ether linkages are the dominant components in both softwood and hardwood lignins<sup>1</sup>, the cleavage of these bonds contributes significantly to lignin degradation. Hence the positive effects of ionizing radiation in pulping<sup>2</sup> may be related to the ease of fragmentation of lignin. A better understanding of these fragmentation processes may also provide possibilities for obtaining lower molecular weight products from lignin.

## 2. CONCLUSIONS

- i) Pulse radiolysis, HPLC, NMR and UV data are consistent with the mechanistic schemes shown in Figs. 6 and 11
- ii) Under oxidative conditions a hydroxylation-cleavage mechanism dominates at acidic and neutral pH while fragmentation also becomes significant at high pH
- iii) A fragmentation pathway, yielding dimethoxyacetophenone (DMAP) and guaiacol predominates under reducing conditions. These products are fairly stable to further radiolysis
- iv) In alkaline medium, under both oxidizing and reducing conditions, radiolytic decomposition of **I** is significant despite the absence of a phenolic group

## 3. EXPERIMENTAL

All chemicals were of high research-grade purity, obtained from commercial sources and were used without further purification. <sup>1</sup>H NMR was done with a Bruker Aspect 3000 and Bruker AMX 500 instruments operating at 300 and 500 MHz respectively, referenced to CHCl<sub>3</sub> (at the Chemistry Department, University of Manitoba, Manitoba, Canada). GC-MS experiments were conducted on a Hewlett Packard Instrument, GC model 5890 and MS model 5970 with an HP-5 column, 12 m, and a linear velocity of 50 cm/s. UV spectra were recorded using a Hewlett Packard 8451A spectrophotometer.

Compound **I** was synthesized in 64% yield according to published procedures (m.p. 90 to 91°C, lit. 85 to 86°C<sup>21</sup>). <sup>1</sup>H NMR and GC-MS data confirmed the structure.

### 3.1 Pulse Radiolysis Experiments

Pulse radiolysis was conducted at the Institute of Isotopes of the Hungarian Academy of Sciences, Budapest, Hungary, using a 4 MeV linear electron accelerator with 80 ns or 2.6  $\mu$ s pulse width. Dosimetry was carried out using 10 mM KSCN. The dose per pulse value was varied between 5 and 20 Gy for the 80 ns pulses, and between 8 and 100 Gy for the 2.6  $\mu$ s pulses. Absorbances (*A*) were converted to G<sub>rx</sub> values by the following formula:



$$G\epsilon_{\lambda} = \frac{A}{D d} - 9.6 \cdot 10^6 (100 \text{ eV})^{-1}$$

where  $D$  is the dose (Gy),  $d$  the density of the solution ( $\text{g cm}^{-3}$ ), and  $G$  the number of species formed, decayed or transformed per 100 eV absorbed. The time-dependent concentration of the optically absorbing species,  $i$ ,  $\epsilon$ , the transient absorption, was measured by an Applied Photophysics K-347 kinetic spectrophotometer. On the oscilloscope screen the time-dependence of the light intensity ( $\Delta I(t)$ , mV) passing through the cell appears in mV units. On the basis of the light intensity before the electron pulse ( $I_0$ , mV) the time dependent light intensity is calculated by the data acquisition system

$$\text{OD}_{\lambda} = \log \frac{I_0}{I_0 - \Delta I(t)} = \epsilon_{\lambda} c l$$

where  $\epsilon_{\lambda}$  is the extinction coefficient at  $\lambda$  wavelength,  $c$  the concentration of absorbing species, and  $l$  the cell light path. Three storage oscilloscopes (100, & 60 MHz Phillips, 100 MHz Iwatsu Ts 6123), coupled with an IBM compatible computer were used for data acquisition<sup>22</sup>

The decay of transient species usually followed second order kinetics and fittings were made using the standard formula

$$\frac{1}{G \epsilon_{\lambda}} = \frac{1}{(G \epsilon_{\lambda})_0} + \frac{2k}{\epsilon_{\lambda}} t$$

where  $2k$  is the second order rate parameter of the self-termination reaction of the intermediates

Experiments were performed on solutions of **I** (0.1 mM) in double-distilled water deaerated using  $\text{N}_2$  or saturated with  $\text{N}_2\text{O}$ . Solubilization of **I** was enhanced by ultrasonication. The pH was adjusted with 7%  $\text{HClO}_4$  or 0.1 M NaOH. Hydrated electrons ( $e_{\text{aq}}^-$ ) were generated in  $\text{N}_2$ -saturated solutions containing 1.0 M *t*-butanol which scavenges hydroxyl radicals and also increases the solubility of **I**. Azide radicals produced by irradiation of solutions containing 100 mM  $\text{NaN}_3$  were used to oxidize phenolic type aromatic substrates.

### 3.2 Steady State Radiolysis

Solutions were prepared as mentioned above. Irradiations were performed in an AECL Gammacell 220. HPLC was performed using a Beckman model 344 pump upgraded with a data acquisition system run by a Nelson Analytical Model 2600 Chromatographic Software system loaded on an IBM PC. Samples were injected onto a Waters Nova-Pak  $\text{C}_{18}$  cartridge (8 x 100 mm) eluted isocratically with MeOH:Water (1:1) at 0.5 mL/min. The eluent was monitored at 274 nm. All HPLC discussed in this paper was performed using these conditions.

HPLC peaks 1, 3, 5, 8, and 9 were identified by comparison of retention times with authentic compounds and also by spiking irradiated solutions of **I** with authentic compounds prior to HPLC. The radiolytic product mixtures seen as peaks 10, and 11, were isolated by repeated HPLC. The collected samples were pooled, concentrated, and analysed by  $^1\text{H}$  NMR. Both 10 and 11 showed complex spectra suggesting a mixture of demethoxylated products. A signal at 3.8 ppm, characteristic of  $\text{OCH}_3$ , was absent indicating complete demethoxylation of all  $\text{OCH}_3$  groups. The exact position of hydroxyl groups attached to the aromatic rings could not be determined due to overlap and signal broadening stemming from the mixture of products.

For indirect characterization of 10, and 11, Compound **I** (3 mg) was refluxed in 5 mL 57% HI for 10 min to produce demethoxylated products. Peaks 10' and 11', corresponding in retention time to peaks 10, and 11 from irradiated samples of

I, were separated by HPLC, collected, and concentrated. The  $^1\text{H}$  NMR of 10' and 11' showed complex spectra of aromatic and aliphatic peaks similar to those found for peaks 10 and 11 from irradiated I. A peak at 5.2 ppm, characteristic of  $\text{CH}_2$ , indicated that the  $\beta$ -aryl ether bond was not fragmented. Integration of signals at 3.8 to 4.0 ppm, (characteristic of  $\text{O-CH}_3$ ) afforded a reduced ratio of  $\text{OCH}_3$  to  $\text{CH}_2$ , indicating a mixture of partially demethoxylated compounds.

To identify the radiolysis products in peak 12, compound I was hydroxylated by standard synthetic techniques. The reaction mixture was analyzed by HPLC. Material corresponding in retention time to peak 12 from irradiated samples was collected by repeated injections and concentrated.  $^1\text{H}$  NMR showed multiple hydroxylation on both rings, with the methylene group and methoxy groups left intact.

#### 4 REFERENCES

- 1 Rydholm, S A , "Pulping Processes," R E Krieger Publishing Company, Malabar, Florida, 1985 186-218
- 2 Granfeldt, T , Jackson, M , Iverson, S L , Free, D , and Chuaqui, C , *Tappi J* 1992 **75** 175-182
- 3 Chuaqui, C , Whitehouse, R , Merritt, J , Iverson, S L , Free, D , Jackson, M , and Granfeldt, T , "The Effects of Electron Beam Treatment on Wood Components" *Proceedings of Tappi Pulping Conference* Orlando, Florida, 1991 Book 2 573
- 4 Chuaqui, C , Merritt, J , Whitehouse, R , Singh, R , and Iverson, S L , "Selected Aspects of the Chemistry and Radiation Chemistry of Lignin" *Proceedings of the Seventh Tihany Symposium on Radiation Chemistry* Hungarian Chemical Society, Dobó, J , Nyikos, L , and Schiller, R , (Eds ), Budapest, 1991 513-519
- 5 Sarkanen, K V , "Precursors and their Polymerization" *Lignins, Occurrence Formation, Structure and Reactions* Sarkanen, K V , and Ludwig, C H , (Eds ) John Wiley & Sons, New York, 1971 95-163
- 6 Adler, E , *Wood Sci Technol* 1977 **11** 169-218
- 7 Shimada, M , and Gold, M H , *Arch Microbiol* 1983 **134** 299-302
- 8 DiCosimo, R D , and Szabo, H-C , *J Org Chem* 1988 **53** 1673-1679
- 9 Dimmel, D R , and Schuller, L F , *J Wood Chem Technol* 1986 **6** 345-65
- 10 Land, E J , and Ebert, M , *Trans Faraday Soc* 1967 **63** 1181-1190
- 11 Burggraf, L W , and Firestone, R F , *J Phys Chem* 1974 **78** 508-514
- 12 Armstrong, R C , and Swallow, A J , *Radiat Res* 1969 **40** 563-579
- 13 O' Neill, P , Steenken, S , Schulte-Frohlinde, D , *J Phys Chem* 1975 **79** 2773-2779
- 14 Steenken, S , and O' Neill, P , *J Phys Chem* 1977 **81** 505-508
- 15 Christensen, H C , Sehested, K , and Hart, E J , *J Phys Chem* 1973 **77** 983-987
- 16 Hayon, E , Ibata, T , Lichtin, N N , and Simic, M , *J Phys Chem* 1972 **76** 2072-2078
- 17 Lundquist, K , and Lundgren, R , *Acta Chem Scand* 1972 **26** 2005-2023
- 18 Ciaramitaro, D , and Steelink, C , *Photochemistry* 1975 **14** 543-552
- 19 Sarkanen, K , and Hoo, L H , *J Wood Chem* 1981 **1** 11-27
- 20 Lundquist, K , *Appl Polym Symp* (Proc Cellul Conf 8th, 1975 Vol 3) 1976 **28** 1393-1407
- 21 Adler, E , Lundgren, B O , and Saeden, U , *Svensk Papperstudning*, 1952 **55** 245-254
- 22 Foldiák, G , Hargittai, P , Kaszanyiczki, L , and Wojnarovits, L , "Radioanalytical and Nuclear Chemistry", Article (1988) **1** 9